# REVIEW

# The role of m6A RNA methylation in cancer metabolism

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# Abstract

Metabolic reprogramming is one of the main characteristics of malignant tumors, which is due to the flexible changes of cell metabolism that can meet the needs of cell growth and maintain the homeostasis of tissue environments. Cancer cells can obtain metabolic adaptation through a variety of endogenous and exogenous signaling pathways, which can not only promote the growth of malignant cancer cells, but also start the transformation process of cells to adapt to tumor microenvironment. Studies show that m6A RNA methylation is widely involved in the metabolic recombination of tumor cells. In eukaryotes, m6A methylation is the most abundant modification in mRNA, which is involved in almost all the RNA cycle stages, including regulation the transcription, maturation, translation, degradation and stability of mRNA. M6A RNA methylation can be involved in the regulation of physiological and pathological processes, including cancer. In this review, we discuss the role of m6A RNA methylation modification plays in tumor metabolism-related molecules and pathways, aiming to show the importance of targeting m6A in regulating tumor metabolism.

Keywords: The m6A, Cancer, Metabolism reprogramming

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modifications have been found [4]. In 1974, a methyl group at the N6 position of adenine was first found in mRNA. is modified base is called m6A, the most abundant internal modification on eukaryotic mRNA.

e average 1000 nucleotides contain 1-2 m6A residues [5]. M6A mainly appears in the RRACH sequences, and is significantly enriched near the stop codon, 3'-UTR and long intron [6]. e basic processes of m6A modification is that it is installed by m6A methyltransferase, removed by m6A demethylase and recognized by m6A reading molecules thus regulating RNA metabolism. More and more evidences show that m6A can influence the expression of target genes, thus regulating a variety of physiological processes, including self-renewal, invasion and proliferation. In the molecular mechanism, m6A is involved in almost all RNA metabolism processes, including translation, degradation, splicing, exporting and folding [7]. In recent years, many studies show that m6A is widely involved in tumor regulation, which further regulates the occurrence and development of tumor by regulating tumor metabolism.

## Cancer metabolism

For many years, the consensus of medical community for tumors is that they are gene-related diseases, almost all cancers are caused by gene changes. Accumulating evidences of cancer phenotypes indicate that all cancers share six biological abilities in the process of multi-step development: continuous proliferation signaling, escape from proliferation inhibitor, resistance to cell death and apoptosis, immortality of replication, induction of angiogenesis, and promotion of invasion and metastasis [8]. However, follow-up studies have shown that cancer has three factors: the target is oncogene or tumor suppressor gene; the abnormal level of m6A in cancer mainly depends on the expression and activity of "writers" and "erasers"; target mRNA is regulated after modification, which is mainly determined by "readers".

At present, the researches on m6A "writers" is the most extensive. e "writers" of m6A are mainly composed of METTL3. METTL14 and their cofactor WTAP ere is an S-adenosylmethionine binding (Table 1). motif in METTL3 and METTL14. ese two genes are located in the nuclear spot together and formed a stable heterodimer complex in the ratio of 1:1 [22]. As a pseudo-methyltransferase, METTL14 plays an important role in stabilizing METTL3 and recognizing target RNAs. As the main regulatory and component molecule of m6A methylation complex, WTAP can help METTL3 and METTL14 locate in the nuclear plaques. In addition, m6A "writers" also include METTL16, KIAA1429 and RBM15. Among them, KIAA1429, as the largest sca old component of m6A methyltransferase complex, is used to regulate 3'-UTR of genes and m6A methylation near the stop codon [23, 24].

Demethylation is mainly acted by FTO and ALKBH5. As a reversible step of m6A methylation, demethylase FTO can regulate fat production and energy homeostasis [25]. ALKBH5 has the highest expression in testis, but lower expression in heart and brain, which can a ect nuclear RNA output, metabolism and gene expression [26].

Besides "writers" and "erasers", another indispensable group in m6A is called "readers". ey can recognize modifications and combine with them. Di erent readers can perform di erent biological functions. e most famous m6A readers are YTHDF family and IGF2BP family [27]. e YTH domains in human cells, including YTHDF1-3 and YTHDC1-2, preferentially bind to family is responsible for recruiting RNA stabilizers to promote mRNA stability, thereby a ecting tumor progression [29].

## The m6A and metabolic signaling pathways

e development of tumor progression needs metabolism reprogramming. In fact, tumor metabolism reprogramming is mainly composed of bioenergy metabolism and biosynthesis metabolism. Bioenergy metabolism is mainly mediated by mitochondrion, while the biosynthesis metabolism is mainly refers to regulation the synthesis of glucose, lipid and amino acid. In addition to the tumor cells, the other cells in tumor microenvironment including endothelial cells, fibroblasts and immune cells also need to undergo metabolic reorganization to meet the needs of promoting tumor progression. Tumor related signaling pathway is a series of enzymatic reaction pathways with various e ects on tumor cells. ousands of genes can regulate tumor progression, but most of them are attributed to the activation or inhibition of tumor-related signaling pathways eventually. For tumor metabolism reprogramming, it is found that di erent metabolism may be concentrated to the same signaling pathway, but play distinct roles. erefore, comprehen-



# The m6A and MAPK

Mitogen activated protein kinase (MAPK) signaling pathway is a critical area to induce tumor, which is involved in a series of cellular physiological activities. MAPK is mainly composed of four subfamilies: extracellular signal regulated kinase (ERK), p38 mitogen activated protein kinase (p38 MAPK), c-Jun N-terminal kinase (JNK) and extracellular signal regulated kinase 5 (ERK5). Among them, RAS-Raf-MEK-ERK signaling pathway has been widely studied and found to be inextricably linked with tumor. Researches show that MAPK signaling participates and plays an essential role in glycolysis. It is reported that as for p38 MAPK participates in aerobic glycolysis through regulating PFKBF3 and Glut2 [37]. Activation of MAPK/Erk signaling can induce c-Myc transcription, thus promoting glycolysis [38].

In pancreatic cancer, METTL3 deficient cancer cells are highly sensitive to gemcitabine, 5-fluorouracil, cisplatin and radiotherapy [39]. is is because METTL3 can regulate the cascade of MAPK, leading to resistance to chemotherapy and radiotherapy [39]. In renal cell carcinoma, ATP receptor P2RX6 may regulate Ca<sup>2+</sup> mediated p-ERK1/2-MMP9 signaling pathway, and increase the invasion ability of RCC cells. METTL14 can increase P2RX6 precursor mRNA splicing, decrease P2RX6 protein translation and decrease P2RX6 mRNA and protein levels through m6A methylation [40]. METTL3 can promote activation of MAPK by promoting the methylation of pri-miR-1246 and targeting SPRED2 in colorectal cancer. Down-regulating SPRED2 further reverses the inhibition of MAPK pathway and promotes the invasion and metastasis of tumor cells [41].

# The m6A and PTEN

PTEN was discovered as a tumor suppressor at first, which mainly through negatively regulating PI3K-Akt signaling pathway. us, PTEN can also participate in tumor progression and glucose-lipid metabolism. PI3K can phosphorylate the metabolite PIP2 to PIP3, while the lipid phosphatase PTEN can dephosphorylate PIP3



Fig. 2

and change it back to PIP2, thus inhibiting its accumulation in cells and terminating the PI3K signaling pathway. In bladder cancer, METTL3 interacts with DGCR8, and actively promotes the maturation of pri-miR-221/222 in a m6A dependent manner, further reduces the expression of PTEN and promotes the proliferation of bladder cancer [42]. As we all known, hepatitis B virus is a direct factor leading to liver cancer. Studies show that hepatitis B virus can significantly increase the m6A modification of PTEN, leading to the instability of PTEN expression, and reduce the expression of PTEN. e expression of PTEN can directly increase the activation of IRF-3, promote the nuclear transcription of IRF-3, thus a ecting the synthesis of interferons, and influence the occurrence and development of liver cancer [43].

## The m6A and AMPK

AMPK, the signaling pathway which looks very similar to MAPK, with the full name is AMP-activated protein kinase. It can regulate energy homeostasis, and involved in a variety of signal transduction pathways. It is one of the central regulators of eukaryotic cells and maintains the smooth operation of cell physiological activities. When the expression of AMP and ADP increases, AMPK is activated. e activated AMPK pathway can a ect metabolism recombination by phosphorylating the substrates and transcriptional regulators. In fact, AMPK and mTOR signaling pathway are also closely related. In the case of nutritional deficiency, AMPK can directly phosphorylate Raptor, block the ability of mTORC1 kinase complex to phosphorylate its substrate, and inhibit cell growth. In breast cancer cells, low-dose SFN can play an anti-tumor role by reducing ATP and AMPK activation pool and stimulating energy stress induced by autophagy, which is caused by promoting DNA hypomethylation and reducing the methylation level of m6A, promoting genetic instability of cancer cells and inhibiting tumor progression [44].

#### The m6A and Wnt

e function of Wnt signaling pathway is mostly in the regulation of embryonic development and cancer. Wntcatenin pathway can lead the accumulation of -catenin in tumor cytoplasm and promote translocation of transcription coactivator/LEF family to the nucleus. Abnormal Wnt signaling is considered to be the driving factor of metabolic changes in glycolysis, glutamine decomposition and adipogenesis, which is crucial for the survival of cancer stem cell population [45]. Glutamine deficiency in cancer stem cells can reduce Wnt signaling activity and induce the increasing phosphorylation of -catenin.

erefore, the regulation of glutamine on stem cell-like cancer cells is partly through the phosphorylation and degradation of -catenin mediated by reactive oxygen species [46].

In hepatoblastoma, the abnormal expression of METTL3 can promote tumor progression, and the modification of m6A in tumor cells generally increases. CTNNB1, as a key component of Wnt signaling, its m6A abundance increases significantly with the expression of METTL3, which enhances the stability of CTNNB1 and then activating Wnt- -Catenin signaling pathway [47]. However, in gastric cancer cells, inhibition of m6A methylation can activate Wnt signaling pathway to promote tumor cell proliferation and invasion, while FTO knockout can reverse these molecular and behaviors changes ese results show that distinct m6A methylation **[32]**. status of Wnt- -catenin signaling might represent opposite role in di erent tumors. In metastatic endometrial carcinoma, FTO can catalyze demethylation 3'-UTR region of HOXB13 mRNA, eliminate the recognition e ect of YTHDF2 on the m6A methylation, reduce the attenuation e ect of HOXB13 mRNA, and increasing the expression of HOXB13 can activate Wnt signaling pathway, thereby promoting tumor invasion and metastasis [48]. In pancreatic ductal adenocarcinoma, silencing ALKBH5 can promote malignant biological behaviors of cancer cells, and also promote the drug resistance of cancer cells to chemotherapy. ALKBH5 can increase the mRNA expression of Wnt inhibitor factor 1 (WIF-1) by reducing the m6A modification of WIF-1 3'-UTR, and then regulates Wnt- -catenin signaling. Recovery experiment shows that the up-regulation of WIF-1 by ALKBH5 can be restored by methylation inhibitor DAA [49].

# The m6A and Hedgehog

Hedgehog signaling molecule is a kind of local protein ligand secreted by signal cells, which has a small range of action and its production is strictly controlled by time and space. Hedgehog signaling pathway is responsible for controlling cell fate, proliferation and di erentiation of cells. Studies demonstrate that when aberrant activating the pathway can cause the occurrence and progression of tumor, which through regulating glycolysis and glutaminolysis [50, 51]. However, there are limited reports on the regulation of Hedgehog pathway by m6A methylation in tumors. In prostate cancer cells, silencing METTL3 reduces the m6A modification of Gli1, an important component of Hedgehog pathway, and reduces the expression of Gli1, as well as the expression of downstream of hedgehog pathway, which promotes tumor cell apoptosis [52].

#### The m6A and NF- B

NF- B is one of the well-known tumor-related signaling pathways, which is often shows in form of homodimer or heterodimer, with p65 and p50. However, NF- B can be inactivated in cytoplasm due to the formation of trimer complex by binding with IkB protein. When the upstream factor TNF binds to the receptor on the membrane surface, the receptor conformation changes and then ram face molehat di

upstrtly thi T-5(o6(h)12(w)-10()-5(lyc)8(oly)-5(si)-5(s)-24(, g)-5(lu(, ).5(

M2-like tumor-associated macrophages and regulatory T cells, and weakened the e cacy of PD-1 checkpoint blockers. e METTL3 deficient mice decreases m6A methylation level of SPRED2, which can't be recognized by YTHDF1, thus the reduction of SPRED2 translation leads to enhance NF- B activation and promote tumor progression [59].

#### The m6A and transcription factors

Transcription factors are proteins that regulate gene transcription process. It is an important part in regulating signal pathways, also known as the third messenger.

erefore, comprehensive analysis of the important role of transcription factors in tumor metabolism is a powerful supplement to investigate the role of signaling pathways in tumor metabolic recombination. erefore, it is very necessary to find out the common signaling pathways and transcription factors correlated with metabolic recombination of tumor cells, immune cells and fibroblasts in the tumor microenvironment, so as to understanding the impacts of metabolic recombination on tumor microenvironment as a whole and promote tumor progression.

#### The m6A and HIF-1

HIF-1 is a well-known hypoxia inducible factor, hypoxia and tumor are mutual positive feedback. Firstly, the increasing oxygen consumption caused by the proliferation of tumor cells promotes the hypoxia environment in the tumor microenvironment. On the contrary, the hypoxia microenvironment can promote the proliferation, di erentiation, energy metabolism and drug resistance of tumor cells through various factors and signaling pathways, thus forming a positive feedback loop to promote the malignant progress of tumor. It is one of the important reasons for poor prognosis of cancer patients. HIF-1 and HIF-2 are heterodimeric transcriptional activators, which consist of HIF-1 and HIF-2 subunits regulated by O<sub>2</sub> and HIF-1 subunits constitutively expressed [60]. Due to the role of HIF plays in tumor microenvironment regulation, HIF can widely participate in tumor metabolic reorganization, including glucose metabolism, lipid metabolism and so on. HIF shows an essential role in regulating glycolysis in the development of various cancers, including regulates glycolysis enzymes of HK2, GLUT1, PKM2 and et al. [61, 62]. Studies have shown that a variety of m6A methylases can participate in the regulation of HIF-1 methylation level and expression, and then participate in tumor progression.

In breast cancer, hypoxia induces demethylation of m6A, mainly HIF-1 and HIF-2, which is highly expressed under hypoxia, can activate the gene transcription of ALKBH5, reduce NANOG m6A methylation and stabilize its stability, increase its protein expression, and promote the phenotype of breast cancer stem cells [63] (Supplementary Table 2). It is well known that the activation of autophagy is one of the important ways for cancer cells to survive in hypoxia. In hepatocellular carcinoma, HIF-1 promotes the expression of YTHDF1 by binding to the promoter of YTHDF1, and then combines with ATG2A and ATG14 by m6A modification to promote their translation and malignant progression of cancer [64].

In HCC, the expression of HBXIP (hepatitis B virus X-interacting protein) is up-regulated, which positively regulates the expression of downstream methylase METTL3 and promotes HIF-1 expression, sustain high level of glycolysis, thus promote malignant biological behavior of HCC cells [65]. In endometrial carcinoma, hypoxia and high level of ALKBH5 expression promote the transcription of SOX2 through demethylation, thereby increasing the stem cell-like phenotype of endometrial carcinoma. Knockout HIFs or ALKBH5 can significantly reduce its tumor initiation ability [66].

#### The m6A and FOXM1

FoxM1 is a common transcription factor regulating tumor cells, which is mainly involved in the regulation of cell cycle and is closely related to the abnormal proliferation and division of tumor cells. In fact, FoxM1 has a strong regulatory e ect on the metabolic recombination of tumor cells. Inhibition of FoxM1 expression significantly decreases the activation and expression of GLUT1 and HK2, and then inhibits the aerobic glycolysis and cell proliferation [67]. In gastric cancer, FoxM1 targets transcription and activation of Prx3, promotes stemness and metabolic reprogramming of gastric cancer cells, and increases the expression of mitochondrial fatty acid oxidative phosphorylation rather than glycolysis, which promotes drug resistance of tumor cells [68]. ese studies suggest that FoxM1, as a transcription factor, can target metabolism-related genes and regulate tumor cell progression.

It is found that ALKBH5 is closely related to the expression of FOXM1, regulating the level of FOXM1 methylation and tumor progression in many tumors. In glioblastoma stem cells, m6A demethylase ALKBH5 is highly expressed, which directly targets the new transcripts of FoxM1 through demethylation, thus increasing the expression of FoxM1, and promoting the stemness and proliferation of tumor cells. HuR, as an RNA binding protein, promotes the expression of FoxM1 by binding to unmethylated 3'-UTR, and participates in the regulation of FoxM1 by ALKBH5. However, it is worth noting that FOXM1-AS is a long non-coding RNA of antisense FoxM1, which can promote the interaction

between ALKBH5 and FoxM1 new transcripts [69]. In oral squamous cell carcinoma, DDX3 can directly regulate the expression of ALKBH5, reduce m6A methylation in FoxM1 and NANOG new transcripts, lead to chemotherapy resistance [70].

#### The m6A and P53

At first, p53 was discovered as an oncoprotein antigen. Later, it was thought that p53 is an important oncogene. In fact, p53 is a broad-spectrum tumor suppressor gene. Wild-type p53 can promote cancer cell apoptosis, and its inactivation plays an important role in tumor formation. In malignant tumors, more than 50% of patients will have p53 mutations. Functionally, p53 acts as a transcription factor which can regulate cell cycle initiation. However, the function of p53 is not limited to directly regulation of cell cycle and apoptosis, which also widely involved in the metabolic reorganization of tumor cells through a variety of signaling pathways. In colorectal cancer, wild-type p53 targets miR-143-3p-PDK2 signaling pathway to regulate tumor cell glucose metabolism and influence chemoresistance [71]. In addition, p53 inhibits glucose consumption and NADPH expression by binding with G6PD [72]. Compared to glycolysis, wild-type p53 prefers to promote mitochondrial respiration when regulates cell metabolic diversity, which partly through trans-activation of SCO2, e studies showed that c-Myc plays an important role in regulating cancer metabolism.

As an independent prognostic factor, c-Myc knockout can inhibit the expression of YTHDF1, thereby inhibiting the proliferation and chemoresistance of tumor cells [84]. In lung cancer, miR-338-5p can inhibit the expression of METTL3, thereby decreasing the m6A modification of c-Myc, down-regulating its expression and inhibiting the proliferation, invasion and migration of lung cancer cells [85]. In OSCC, METTL3 targets the 3'-UTR of c-Myc transcript to increase the m6A modification. YTHDF1 cooperates with the METTL3 m6A e ects to enhance the stability of c-Myc, therefore knockout METTL3 can inhibit the malignant progression of tumor cells [86]. In gastric cancer, HDAC3 can accelerate the invasion and migration of gastric cancer cells by targeting FOXA2. FOXA2 can bind to the promoter of m6A eraser FTO and reduce its expression. However, FTO can reduce the methylation of c-Myc in gastric cancer cells and stabilize its expression, thereby a ecting the tumor initiation activity of gastric cancer cells [87]. In addition, the expression of METTL3 in gastric cancer cells can also a ect the carcinogenic function of tumor cells, and its overexpression can promote tumor progression. MCM5 and MCM6 are the molecules targeted by c-Myc, and METTL3 can regulate MCM5 and MCM6 through m6A modification. Knockout METTL3 significantly reduces the m6A levels of MCM5 and MCM6, thus inhibiting carcinogenic of gastric cancer [48]. In the acute myeloid leukemia (AML) carrying t (11q23), t (15;17) or t (8;21), METTL14 is up-regulated, and the down-regulation of METTL14 accompany with myeloid di erentiation. Silencing METTL14 can promote the terminal di erentiation of AML cells, thus inhibit proliferation of AML cells. METTL14 plays a carcinogenic role by modifying the downstream expression of MYB-MYC in an m6Adependent manner. Inhibition METTL14 expression results in a significant decreasing of the half-life of MYB and MYC transcripts, thus inhibiting tumor progression [88].

# The m6A and SRF

SRF (serum response factor) is a member of MADS box transcription factor superfamily. Its expression is highly conserved and participates in many important life activities of cells. Although the research on the relationship between SRF and metabolism reprogramming is very limited, some studies have shown that SRF is involved in high glucose induced epithelial mesenchymal transition or high glucose induced damage to retinal ganglion cells. High glucose stimulates the overexpression of SRF, which promotes the epithelial mesenchymal transition of human peritoneal mesothelial cells induced by high glucose through directly binding to Snail promoter [89]. In tumor cells, SRF can promote cell reprogramming into multifunctional cells [90]. SRF, together with MRTF and TCF, can regulate the migration, invasion, growth and proliferation of tumor cells in a signal and cytoskeleton dependent manner [91]. e synergistic e ect of Myc and RhoA-SRF pathway has a synthetic lethal e ect, which is caused by the insu cient utilization of glutamine, suggesting that Myc and RhoA-SRF have metabolic coordination in maintaining the vitality of cancer cells [92].

IGF2BP1 promotes SRF expression in an m6A dependent manner, enhances SRF dependent transcriptional activity at the post-transcriptional level, and promotes tumor cell proliferation and invasion. e results show that knockout METTL3 and METTL14 down-regulates the expression of SRF in cells, while in the SRF-3'-UTR mutant cancer cells, the deletion of METTL3 and METTL14 don't a ect the expression of SRF. ese results suggest that IGF2BP1 enhances SRF expression through a conserved 3'-UTR and m6A dependent manner, and then regulates tumor progression [93]. However, the researches on regulation of SRF by m6A methylation in tumor are very limited, more researches are needed to clarify the role of m6A methylation in regulating SRF in tumor metabolism.

# The m6A and OCT4

OCT4 is a member of the POU transcription factor family, which has many subtypes. e translated proteins contain a conservative DNA binding domain, namely the POU binding domain, which is involved in regulating the stem cell stemness and di erentiation. In human embryonic stem cells, silencing GLUT3 leads to the decrease of glucose uptake, lactate production and OCT4 expression, suggesting that the self-renewal of human embryonic stem cell hat the self-reTd /eru9(a)-12(s)-6(e )]Tg GLis a memb for maintaining the self-renewal of embryonic stem cells. In addition, Sox2 can act as one of the initial factor of inducing pluripotent stem cells. In ovarian cancer, as a transcription factor, Sox2 directly binds to the promoter of ST6Gal-I to drive transcription and increases in N-glycosylated protein 2-6 linked sialic acid, promotes ST6Gal-I and 2-6-linked sialic acid expression, regulates glucose metabolism in cancer cells [97].

In endometrial cancer, PADI2 (peptide arginine deaminase II) can convert arginine residues to citrulline, participates in the regulation of amino acid metabolism, catalyzes the citrullination of MEK1-R113/189, promotes the phosphorylation of ERK1/2, and activates the expression of IGF2BP1. IGF2BP1 can bind to the m6A site of Sox2 3'-UTR to inhibit its degradation, thus promoting the progression of endometrial cancer [98]. Studies have found that METTL3 plays a role of proto-oncogene in colorectal cancer, promotes the methylation level of Sox2 in a m6A dependent manner, prevents Sox2 degradation through the synergistic e ect of IGF2BP2, and maintains the expression of Sox2, thus promoting the progression of colorectal cancer [99]. In bladder cancer, the over-expression of METTL3 increases m6A methylation status, regulates the expression of AFF4, promotes its transcription by combining with the promoter region of Sox2, and promotes the self-renewal of cancer stem cells [100].

#### The m6A and ETS-1

ETS-1, a member of ETS transcription factor family, has a conserved ETS-DNA binding domain and is a key factor in NK cell di erentiation. In pancreatic cancer cells, silencing ETS-1 reduces the expression of GLUT-1, interferes with glycolysis and reduces glucose utilization and lactate production, reduces the energy produced in the form of ATP, and inhibits the vitality and invasion ability of tumor cells [101]. WTAP is an important part of m6A writer, and its main function is to recruit methylase METTL3 and METTL14. As a transcription activator, ETS1 is regulated by Ras-RAF-MEK-ERK signaling pathway [102]. In HCC, WTAP interacts with RNA binding protein HuR to regulate the transcription inhibition of ETS1 by m6A modification, and then regulates the downstream p21-p27 signal axis to regulate the progression of HCC [103].

# The m6A and Snail

Zinc finger transcription factor superfamily Snail family is very conservative in evolution. e amino acid end of the Snail family members contains evolutionarily conserved SNAG domain, which plays an important role in the regulation of transcription inhibition and participates in embryonic development and tumorigenesis. Snail itself is a highly unstable protein, regulated by comprehensive and complex network signals. In gastric cancer, up-regulation of Snail promotes lactate production and glucose utilization, reduces the expression of FBP1, which is the rate limiting enzyme of gluconeogenesis, plays a positive role in regulating glucose metabolism, promotes glycolysis, and then promotes EMT of tumor cells [104].

In HCC, knockout of METTL3 decreases the expression of Snail and inhibits the progression of HCC. Sumo binding enzyme UBC9 regulates the SUMOylation of METTL3, controls homeostasis and promotes the accumulation of Snail, thus promotes the progression of HCC [105]. On the contrary, it is found that the expression of Snail mRNA decreases in the cells with SUMO mutant of METTL3 [106]. In addition, m6A can trigger the polysome mediated translation of Snail, which is in the CDS region of Snail instead of the 3'-UTR region, promote Snail transcription and participate in regulation EMT phenotype of tumor cells. Meanwhile, m6A reader YTHDF1 synergistically increases the translation of Snail mRNA which mediated by m6A methylation [107].

#### The m6A and non-coding RNA

Non-coding RNA is a kind of non-coding transcripts that do not encode proteins, but can produce non-co0 9.8 304.72439

regulated by m6A methylation in influencing the cancer metabolism pathways need to be further investigated.

# The m6A and metabolic

Glyc abivisis

METTL3; WTAP;

IGF2BP3

METTL3

YTHE

valddd<sup>e</sup>i d<sup>h</sup>ladala

# The m6A and glucose metabolism

Glucose metabolism can be divided into catabolism and anabolism. Its metabolic pathways mainly include anaerobic digestion of glucose, aerobic oxidation, pentose phosphate pathway and glycogen synthesis and decomposition. It is well known that abnormal glucose metabolism is an important part of metabolic reprogramming in tumor cells. Among them, a marker of high invasiveness of cancer cells is energy metabolism, including the increase of glycolytic activity and lactic acid fermentation, namely Warburg e ect<sup>9, 136</sup>. Warburg e ect is a typical feature of abnormal glucose metabolism in tumor. It can enhance glycolysis, glucose uptake and consumption, and then make tumor cells more easily adapt to

Fructose-6-P

Fructorose Biston Bandson 3-phosphoglycerate ¥

PEP

ENO1

PKM2

adverse living environment for malignant proliferation. Study shows that Warburg e ect can promote the release of circ-0072083 in exosomes of resistance glioma cells, which targeting miR-1252-5p to regulate ALKBH5, demethylates NANOG and promotes its expression, further promotes TMZ resistance in glioma cancer cells [117]. In non-small cell lung cancer, METTL3 induces m6A methvlation of LncRNA ABHD11-AS1, enhances the stability of ABHD11-AS1 transcript to increase its expression, and promotes the proliferation of tumor cells and Warburg e ect [120]. In colorectal cancer, the overexpression of m6A reader IMP2 (IGF2BP2) can stabilize ZFAS1/ OLA1 axis, which increases the recruitment of OLA1, ATP hydrolysis and glycolysis, activates Warburg e ect, and enhances cell proliferation and colony formation of cancer cells [121].

HK2 is the first important rate limiting enzyme in glycolysis (Fig. 3). METTL3 targets the 3'-UTR of HK2

synthesis



KTA 41420 TTD CL 1 CTT

IGF Za

IGF2BP

WIAt'

FTO

mRNA. METTL3 recruits the m6A reader YTHDF1 to enhance the stability of HK2, thus promoting the Warburg e ect of cervical cancer [122] (Table 2). Extracellular acidification rate (ECAR) assay shows that METTL3 can significantly promote glycolysis. After METTL3 knockout and YTHDF1 silencing, the half-life of HK2 mRNA is significantly shortened. In conclusion, the combination of METTL3 and YTHDF1 enhances the stability of HK2 [122]. In gastric cancer, m6A writer WTAP can promote the proliferation and glycolysis of tumor cells, and knockout of WTAP can inhibit tumor progression.

e mechanism is that through binding the 3'-UTR of HK2, WTAP can enhance the stability of HK2 mRNA, the carcinogenic e ect of WTAP and its m6A mediate regulation of Warburg e ect in gastric cancer, which provides a new way and target for the treatment of gastric cancer [123].

18F-FDG, or fluorodeoxyglucose, known as "century molecule", can accurately reflect the metabolic level of glucose, is the main imaging agent of PET-CT imaging. Due to the high metabolism of cancer cells and the increasing demand for glucose, most tumor lesions show high uptake after intravenous injection of 18F-FDG, so as to accurately determine the primary and

# Table 2 The relationship between m6A enzymes and cancer metabolism

Classi cation	Cancer Type	M6A Related Enzymes	Biological Behavior Changes	Related Enzymes	O cial Full Name of the Enzymes	References
Glucose metabolism	Cervical cancer	METTL3; YTHDF1	Promote Warburg e ect and glycolysis	HK2	Hexokinase 2	[122]
	Gastric cancer	WTAP	Promote the prolif- eration and glycolysis of tumor cells	HK2	Hexokinase 2	[123]
	Colorectal cancer	METTL3; IGF2BP2; IGF2BP3	Activate glycolysis pathways	HK2; SLC2A1	Hexokinase 2; Solute carrier family 2 mem- ber 1	[124]
	Gastric cancer	KIAA1429	Promote aerobic glycolysis to promote tumor progression	GLUT1	Glucose transporter 1	[126]
	Pancreatic ductal adenocarcinoma	YTHDC1	Attenuation Warburg e ect to inhibit tumor progression	SLC2A1; HK1	Solute carrier family 2 member 1; Hexoki- nase 1	[127]
	Bladder cancer	ALKHB5	Regulate glyco- lysis pathways and glucose absorption, lactate and ATP production	CK2	Casein Kinase 2	[128]
	Hepatocellular carci- noma	FTO	Accumulate gly- colysis into anabolic pathways	PKM2	Pyruvate Kinase M2	[129]
	Colorectal cancer	IGF2BP2	Promote glycolysis	c-MYC	MYC proto-oncogene	[131]
	Lung adenocarci- noma	FTO; YTHDF1	Promote tumor glycolysis and tumo- rigenesis	c-MYC	MYC proto-oncogene	[132]
	GBM	FTO	Promote aerobic gly- colysis and promote chemoresistance	PDK1	Pyruvate dehydroge- nase kinase 1	[133]
	Cervical cancer and Liver cancer	YTHDF1; IGF2BP3	Promote glycolysis and ATP generation	PDK4	Pyruvate dehydroge- nase kinase 4	[134]
	Breast cancer	WTAP	Promote glycolysis and promote tumor progression	ENO1	Enolase 1	[135]
	Renal cell carcinoma	METTL14	Promote tumor cell distal lung metastasis	BPTF	Bromodomain PHD finger transcription factor	[136]
Fatty acid metabolism	Hepatocellular carci- noma	FTO; YTHDF2	Influence the content of adipogenic enzymes and intra- cellular lipids	FASN	Fatty acid synthase	[141]
	Esophageal cancer	HNRNPA2B1	Promote cellular lipid accumulation to promote tumor progression	ACLY; ACC1	ATP citrate lyase; Acetyl-CoA carboxy- lase	[143]
Amino acid metabo- lism	Renal clear cell carcinoma	FTO	Synthetic death with VHL and activate VEGF and PDGF	SLC1A5	Solute carrier family 1 member 5	[146]
	Colorectal cancer	IGF2BP1	Promote the tumori- genesis	MYC	MYC proto-oncogene	[147]

# Table 2 (continued)

Classi cation	Cancer Type	M6A Related Enzymes	Biological Behavior Changes	Related Enzymes	O cial Full Name of the Enzymes	References
Mitochondrial metabolism	Renal clear cell carcinoma	METTL3	Regulate one carbon metabolism and aerobic glycolysis of tumor cells	MTHFD2; HIF-2	Methylenetetrahydro- folate dehydrogenase 2; Hypoxia inducible factor-2	[151]
	Renal clear cell carcinoma	FTO	Regulate mitochon- drial activity and pro- mote oxidative stress and ROS production	PGC-1	PPARG coactivator 1	[153]
	Breast cancer	METTL3	Inhibit apoptosis of mitochondrial, attenuate resistance to tamoxifen	AK4	Adenylate kinase 4	[154]

## The m6A and fatty acid metabolism

Lipid metabolism is a complex biochemical reaction in organism, including digestion, absorption, synthesis and decomposition of fat under di erent types of enzymes, which is essential for maintaining cell homeostasis. In cancer, because of the high demand for nutrients, tumor cells often regulate and utilize lipid metabolism to maintain their own proliferation, survival, invasion and metastasis. De novo synthesis of fatty acids is an important metabolic marker in cancer. Enhanced adipogenesis provides an important substance and energy source for tumor growth [137]. Fatty acid oxidation is an important source of cellular energy. Inhibition of fatty acid oxidation can inhibit the growth of tumor cells [138]. Up-regulation of FAO may maximize the production of ATP, reduce intracellular ROS and protect cancer cells from death [139]. Lipid transformation induces the activation key fatty acid synthesis enzymes in tumor cells, such as acetyl CoA carboxylase (ACC) and fatty acid synthase (FAS), and then promotes the proliferation and survival of cancer cells [140].

In hepatoma cells, FTO knockout significantly reduces the content of new adipogenic enzymes and intracellular lipids. is is because FTO knockout makes YTHDF2 play its recognition function, significantly increases the level of FASN m6A, decreases the stability and expression of FASN, and significantly decreases the protein levels of acetyl CoA carboxylase ACC and ATP citrate lyase, thus inhibiting the formation of new fat, leads to insucient lipid accumulation and induces apoptosis [141].

Carnitine palmitoyltransferase 1B (CPT1B) is the rate limiting enzyme of fatty acid oxidation. High m6A methylation level in drug-resistant cells can trigger the splicing of ESRRG precurser, increases the expression of CPT1B and induces up-regulation of ERR in chemoresistant cells, which can promote fatty acid oxidation of tumor cells and enhance chemoresistance of tumor cells [142]. In esophageal cancer, m6A reader HNRNPA2B1 can upregulate fatty acid metabolism-related genes ACLY and ACC1, and thus promote cellular lipid accumulation, which can further promote tumor progression, including proliferation, migration and invasion of cancer cells [143].

## The m6A and amino acid metabolism

Amino acids are produced by the hydrolysis of proteins.

e metabolism of amino acids in the body includes two main aspects: one is the synthesis of proteins, peptides and other nitrogen-containing substances needed for their own synthesis; the other is the decomposition amino acids through deamination, transamination functions to produce -ketoacid, CO2, etc. Among them,

-ketoacids can release energy through TCA oxidation [144]. Serine, glycine and other nonessential amino acids are closely related to the occurrence and development of cancer, so inhibiting the activity of these nonessential amino acids can be used as a potential means of cancer treatment. TCA is not only the final metabolic pathway of the three nutrients, but also the hub of carbohydrate, lipid and amino acid metabolism. In order to maintain a functional TCA cycle, cancer cells usually rely on the elevation of glutamatelyase. erefore, glutamine decomposition is another important feature of tumor energy metabolism [145].

VHL protein is HIF family substrate recognition site, targeting HIF family can degrade ubiquitin mediated proteasome. In renal clear cell carcinoma, the lack of tumor suppressor gene VHL is a significant sign. VHL and FTO have the function of synthetic death. e inactivation of VHL leads to the structural activation of VEGF and PDGF, which can target downstream glutamine transporter SLC1A5, promotes the metabolic reprogramming of VHL deficient renal cancer cells, selectively reduces the growth and survival of VHL deficient renal cancer cells [146].

Long non-coding RNA Linc00266-1 can encode a peptide composed of 71 amino acids, which is called RNA binding regulatory peptide (RBRP). As long as the peptide mainly interacts with RNA binding proteins including IGF2BP1, Linc00266-1, as a subunit regulated by m6A reader, enhances the recognition of c-Myc m6A site by IGF2BP1, increases the mRNA stability and expression level of c-Myc and promotes the tumorigenesis [147].

Histone modifications have huge e ects on gene expression. Huang et al. reported that histone H3 trimethylation at lysine 36 (H3K36me3) significantly enriched m6A modification, which can be recognized and bounded by METTL14. In the embryonic stem cells of mouse, the knockdown of METTL14 and depletion of H3K36me3 significantly reduces m6A abundance, and increases the stemness of cells [148]. ALKBH1 can also works as an m6A demethylation enzyme. In lung cancer, ALKBH1 is up-regulated, due to its function of demethylation the essential residues Y184, H231, D233, H287, R338 and R344, it can significantly promote tumor migration and invasion of cancer cells [149].

# The m6A and mitochondrial metabolism

As the energy factory of cells, mitochondria produce ATP to cells by burning glucose, lipids and amino acids, which can complete various life activities for cell functions. However, in recent years, studies have found that there are many correlations between mitochondrial metabolism and tumorigenesis. At the same time, because mitochondria are the metabolic center of cells, and tumor cells have the characteristics of abnormal metabolism, the development of compounds targeting mitochondria has become a new direction of anti-tumor research. Although the Warberg e ect has been verified in all kinds of tumorigenesis, it has been found that oxidative phosphorylation and mitochondria dependent energy synthesis are the key processes to maintain the stemness of some tumor cells in recent years [150]. Many studies have found that the growth of tumor cells can be regulated through mitochondrial metabolism reprogram.

One carbon metabolism, including folate cycle, methionine cycle and sulfur transfer pathway, plays a key role in a series of metabolic processes required for tumor cell survival and growth. Methylenetetrahydrofolate dehydrogenase 2 (MTHFD2) is a mitochondrial enzyme involved in one carbon metabolism, which regulates HIF-2 transcriptomic mechanism thus influences the progression of RCC. Although MTHFD2 has not been identified as an m6A methylase by definition, MTHFD2 expression is significantly increased in renal cell carcinoma and is involved in regulation the overall level of m6A methylation, especially HIF-2 m6A methylation, promotes HIF-2 expression, thus promotes aerobic glycolysis of tumor cells and tumor progression. One carbon metabolism is associated with HIF-2 dependent metabolic reprogramming combined with the RNA methylation modification. MTHFD2 can regulate mRNA methylation and specifically increase the methylation level of METTL3 dependent HIF-2 . HIF-2 in turn can bind to the promoter region of MTHFD2 gene, and its overexpression increases the level of MTHFD2, thus forming a positive feed-forward loop to promote metabolic recombination and tumor progression [151].

Meclofenac is a non-steroidal anti-inflammatory drug, which is mainly used in the treatment of arthritis, analgesia and dysmenorrhea. However, recent studies have found that meclofenac can also be used as a highly selective FTO inhibitor to reduce ROS accumulation and apoptosis, and participates in the regulation of mitochondrial function [152]. FTO plays an antitumor role in renal clear cell carcinoma. Chronic mitochondrial dysfunction can lead to the loss phenotype of VHL (von Hippel Lindau), the most common mutated tumor suppressor gene in renal cell carcinoma. Accumulation of metabolites during mitochondrial dysfunction can inhibit the degradation of VHL dependent HIF- , pseudo hypoxia state similar to the loss of VHL

#### Implication in chemotherapy

Chemotherapy is one of the most e ective means to treat cancer. Together with surgery and radiotherapy, it is called the three major treatment means of cancer. e purpose of chemotherapy is to cure tumor, delay tumor metastasis, alleviate tumor symptoms and minimize toxic reactions.

Studies demonstrate that the m6A modification of genes show the relevance to chemotherapy in cancer. In pancreatic cancer, METTL3-depleted cancer cells show higher sensitivity to gemcitabine, 5-FU and cisplatin [**39**] (Table **3**). is result shows that m6A methylation regulated cancer can play an important role in chemotherapy. In non-small cell lung cancer, in order to adapt hypoxia microenvironment, YTHDF1 is down-regulated, which further causes the resistance to cisplatin through regulating the translation of CDK2, CDK4 and cyclin D1 [155]. However, in colorectal cancer, YTHDF1 is significantly up-regulated, which reduces the sensitivity to cisus, YTHDF1 expression shows totally di erent platin. expression models in distinct cancer. In fact, YTHDF1 can bind to the 3'-UTR of GLS1, and promote the protein synthesis of GLS1. us, YTHDF1 promotes cisplatin resistance through reprogramming GLS1-glutamine metabolism in colorectal cancer [156]. In leukemia, the resistance to tyrosine kinase inhibitor (TKI) is dependent on over-expression of FTO [157]. PARP inhibitor is an e ectively targeting therapeutic drug in ovarian cancer, which mainly used to platinum sensitive recurrent ovarian cancer patients, thus the resistance to PARP inhibitor is a big challenge for ovarian cancer patients. e m6A methylation level can contribute to PPAR inhibitors resistance to the BRCA deficient cancer cells through up-regulating Wnt/ -catenin pathway via stabilizing FZD10. e increases of FZD10 m6A level, which mediated by down-regulation of FTO and ALKBH5, promotes PARP inhibitor resistance, which is due to the increase of homologous recombination activity [158]. In pancreatic ductal adenocarcinoma (PDAC), ALKBH5 works as a tumor suppressive gene, which shows more chemosensitizing to gemcitabine. In addition, e silencing of ALKBH5 significantly promotes proliferation, migration and metastasis of cancer cells [49].

# Implication in immunotherapy

PD-1 is a star immunosuppressive molecule, which can down-regulate the response of the immune system, and regulates the immune system by inhibiting the activity of T cells, so as to promote immune tolerance. In tumors, PD-1 can help tumor cells escape from the immune system. At present, there are many monoclonal antibodies targeting PD-1 on the market and used in clinic. However, in some conditions, the response of tumor patients to anti-PD-1 therapy is limited, so it is necessary to combine with other targeted drugs. In colorectal cancer and melanoma, knockout of METTL3 and METTL14 inhibits m6A modification, stabilizes STAT1 and IRF1 mRNA expression, and promotes IFN -Stat1-Irf1 signaling transduction through the synergistic e ects of YTHDF2, which increases the expression of CD8<sup>+</sup> T cells, and increases the secretion of IFN-, CXCL9 and CXCL10, enhances the response of pMMR-MSI-L to PD-1 treatment [159].

In human acute myeloid leukemia (AML), FTO promotes the occurrence of leukemia. FB23 and FB23-2, as the inhibitors of FTO, can directly bind with FTO to

Classi cation	Cancer Type	M6A Related Enzymes	Biological Behavior Changes	References
Chemotherapy	Pancreatic cancer	METTL3	Decrease sensitivity to gemcitabine, 5-FU and cisplatin	[39]
	Non-small cell lung cancer	YTHDF1	Promote chemosensitivity to cisplatin	[155]
	Colorectal cancer	YTHDF1	Promote cisplatin resistance through reprogram GLS1-glutamine metabolism	[156]
	Leukemia	FTO	Promote resistance to tyrosine kinase inhibitor (TKI)	[157]
	Ovarian cancer	FTO; ALKBH5	TO; ALKBH5 FOR the sensitivity to P(O; ALKBH5)]Tt 9.8 0 0 c	
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proliferator activated receptor; PRR5: Proline rich 5; PTEN: Phosphatase and tensin homolog; RBM15: RNA binding motif protein 15; RBRP: RNA binding regulatory peptide; ROS: Reactive oxygen species; SLC1A5: Solute carrier family 1 member 5; SOX2: Sex determining region Y-box 2; SPRED2: Sprouty-related EVH1 domain containing 2; SRF: Serum response factor; STAT: Signal transducer and activator of transcription; SUMO: Small ubiquitin-like modifier; TAZ: Tafazzin; TCA: Tricarboxylic acid; TLR4: Toll-like receptor 4; TMZ: Temozolo-mide; TNF: Tumor necrosis factor; TRIM25: Tripartite motif containing 25; UBC9: Ubiquitin-conjugating enzyme E2I; UTR: Untranslated region; VEGF: Vascular endothelial growth factor; VHL: Von Hippel-Lindau tumor suppressor; VIRMA: Vir like m6A methyltransferase associated; WIF-1: Wnt inhibitor factor-1; WTAP: Wilms tumor 1 associated protein; XIST: X-inactive specific transcript; YAP: Yes-associated protein; YTHDC: YTH domain-containing; YTHDF: YTH domain-containing family protein.

# Supplementary Information

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Additional le 1: Table 1. The metabolism-related pathways in cancer with m6A enzymes. Table 2. Metabolism-related transcription factors with m6A enzymes. Table 3. The metabolism-related non-coding RNA in cancer with m6A enzymes.

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#### Authors' contributions

A.Y. and D.H. conceived the review. A.Y. wrote the first version of the manuscript. D.H. revised the manuscript. All the authors approved the final version of the manuscript.

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#### Availability of data and materials

All the data obtained and/or analyzed during the current study were available from the corresponding authors on reasonable request.

#### Declarations

# Ethics approval and consent to participate

Not applicable.

# Consent for publication

All authors give consent for the publication of manuscript in Molecular Cancer.

#### **Competing interests**

The authors declare that there is no potential competing interest.

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#### References

- Lunt SY, Vander Heiden MG. Aerobic glycolysis: meeting the metabolic requirements of cell proliferation. Annu Rev Cell Dev Biol. 2011;27:441–64.
- 2. Warburg O. On the origin of cancer cells. Science. 1956;123:309-14.
- Scharping NE, Rivadeneira DB, Menk AV, Vignali PDA, Ford BR, Rittenhouse NL, Peralta R, Wang Y, Wang Y, DePeaux K. Mitochondrial stress induced by continuous stimulation under hypoxia rapidly drives T cell exhaustion. Nat Immunol. 2021;22:205–15.
- Boccaletto P, Machnicka MA, Purta E, Piatkowski P, Baginski B, Wirecki TK, de Crécy-Lagard V, Ross R, Limbach PA, Kotter A, et al. MODOMICS:

a database of RNA modification pathways. Nucleic Acids Res. 2018;46:D303-d307 (2017 update).

- Krug RM, Morgan MA, Shatkin AJ. Influenza viral mRNA contains internal N6-methyladenosine and 5'-terminal 7-methylguanosine in cap structures. J Virol. 1976;20:45–53.
- Meyer KD, Saletore Y, Zumbo P, Elemento O, Mason CE, Ja rey SR. Comprehensive analysis of mRNA methylation reveals enrichment in 3' UTRs and near stop codons. Cell. 2012;149:1635–46.
- Liu O, Gregory RI. RNAmod: an integrated system for the annotation of mRNA modifications. Nucleic Acids Res. 2019;47:W548-w555.
- Hanahan D, Weinberg RA. The hallmarks of cancer. Cell. 2000;100:57–70.
   Hanahan D, Weinberg RA. Hallmarks of cancer: the next generation.
- Cell. 2011;144:646–74.
- Guerra F, Arbini AA, Moro L. Mitochondria and cancer chemoresistance. Biochim Biophys Acta Bioenerg. 2017;1858:686–99.
- 11. Lin W, Zhou Q, Wang CQ, Zhu L, Bi C, Zhang S, Wang X, Jin H. LncRNAs regulate metabolism in cancer. Int J Biol Sci. 2020;16:1194–206.
- Pavlova NN, Thompson CB. The Emerging Hallmarks of Cancer Metabolism. Cell Metab. 2016;23:27–47.
- Pavlides S, Whitaker-Menezes D, Castello-Cros R, Flomenberg N, Witkiewicz AK, Frank PG, Casimiro MC, Wang C, Fortina P, Addya S, et al. The reverse Warburg e ect: aerobic glycolysis in cancer associated fibroblasts and the tumor stroma. Cell Cycle. 2009;8:3984–4001.
- 14. Wu H, Ying M, Hu X. Lactic acidosis switches cancer cells from aerobic glycolysis back to dominant oxidative phosphorylation. Oncotarget. 2016;7:40621–9.

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methyltransfocellular carcinoma. Nat Commun. 2018;9:1572.

- Wang T, Kong S, Tao M, Ju S. The potential role of RNA N6-methyladenosine in Cancer progression. Mol Cancer. 2020;19:88.
- Mirabilii S, Ricciardi MR, Tafuri A. mTOR Retabolism in Hematologic Malignancies. Cells. 2020;9:404.
- Zhang C, Zhang M, GW, Lin X, Gao J Redicts malignant phenotypes and augmented Wnt/PI3Knaling in gastric cancer. Cancer Med. 2019;8:4766–81.
- Zhang H, Zhang P, La X. m(6)A methyltransfTTL3 promotes retinoblastoma progression via PI3K/AKT/mTOR pathway. J Cell Mol Med. 2020;24:12368–78.
- Liu Jckert MA. m(6)A mRNA methylation regulates AKT activity to promote the prolifigenicity of endometrial cancer. Nat Cell Biol. 2018;20:1074–83.
- Wang X, Zhao BS, Ree IA, L, Ma H, Weng X, Chen K, Shi H, He C. N(6)-methyladenosine Modulates Messenger RNA Translation E ciency. Cell. 2015;161:1388–99.
- Feng ST, Qiu GO, Yang LH, Feng LH, Fan X, R Huang KD, Chen Y. Omeprazole improves chemosensitivity of gastric cancer cells by m6A demethylase FTOed activation of mTORC1 and DDIT3 upregulation. Biosci R. 2021;41(1):BSR20200842.
- Wang F, Qi XM, Wertz R, Mortensen M. p38 MAPK Is Essential ferobic Glycolysis and Pancreatic Tumorigenesis. Cancer R. 2020;80:3251–64.
- Wei X, Zhang Y, Li C, Ai K, Li K, Li H, Yang JThe evolutionarily conserved MAPK/Erk signaling promotes ancestral T-cell immunity in fish via c-Myc-mediated glycolysis. J Biol Chem. 2020;295:3000–16.
- Taketo K, Konno M, Asai A, Koseki JToratani M, Satoh T, Doki Y, Mori M, Ishii H, Ogawa K. The epitranscriptome m6A writer METTL3 promotes chemo- and radioresistance in pancreatic cancer cells. I. 2018;52:621–9.
- G, Zhang JY, Xu Y, Ma JY, Zheng JW, Xue W. The m(6)A-suppressed P2RX6 activation promotes renal cancer cells migration and invasion through ATP-induced Ca(2+) influx modulating ERK1/2 phosphorylation and MMP9 signaling pathway. J Exp Clin Cancer R. 2019;38:233.
- Peng W, Li J, Yang P, Qian W, Ji D, Wang Q, Zhang Z, Tang JY. Upregulated METTL3 promotes metastasis of colorectal Cancer via miR-1246/SPRED2/MAPK signaling pathway. J Exp Clin Cancer R. 2019;38:393.
- Han JWang JZ, Yang X, Yu H, Zhou R, LYuan WB, L L, et al. METTL3 promote tumor proliferation of bladder cancer by accelerating pri-miR221/222 maturation in m6A-dependent manner. Mol Cancer. 2019;18:110.
- K, IV-leased N6 Methyladenosine Modification of PTEN RNA A ects le Immunity and Contributes to HCC. Hepatology. 2020;73(2):533–47.
- Lwinska A, Adamczyk-ochala Jeregowska A, Wnuk M. Sulf-lell Cycle Arrest and Se accompanied by DNA Hypomethylation and Changes in microRNA Profile in Breast Cancer Cells. Theranostics. 2017;7:3461–77.
- El-Sahli S, Xie Y, Wang L, Liu S. Wht Signaling in Cancer Metabolism. Cancers (Basel). 2019;11:904.
- Liao J, Hou G, Shao J, Yang J, LW, Wen S, Hu Y, Huang P. Rem-like cancer cells by glutamine through -catenin pathway mediated by redox signaling. Mol Cancer. 2017;16:51.
- Liu L, Wang J, Sun G, Wu Q, Ma Ju M, et al. m(6)A mRNA methylation regulates CTNNB1 to promote the prolifoblastoma. Mol Cancer. 2019;18:188.
- Zhang L, Wan Y, Zhang Z, Jiang Y, Lang J: FTO demethylates m6A modifications in HOXB13 mRNA and promotes endometrial cancer metastasis by activating the WNT signalling pathway. 2020:1–14.
- Tang B, Yang Y, Kang M, Wang Y, Wang Y, Bi Y, He S, Shimamoto F. m(6) A demethylase ALKBH5 inhibits pancreatic cancer tumorigenesis by decreasing WIF-1 RNA methylation and mediating Wnt signaling. Mol Cancer. 2020;19:3.
- 50. SpannI S, Buhl T, Nellas I, Zeidan SA, Iyer KV, Khaliullina H, S Glycolysis regulates Hedgehog signalling via the plasma membrane potential. EMBO J

- Du K, Hyun Jremont RT, Choi SS, Michelotti GA, Swiderska-Syn M, Dalton GD, Thelen E, Rizi BS, Jung Y, Diehl AM. Hedgehog-YAP Signaling Pathway Res Glutaminolysis to Control Activation of Hepatic Stellate Cells. Gastroenterology. 2018;154:1465-1479.e1413.
- Cai JYang F. RNA m(6)A Methyltransferase METTL3 Promotes The Growth Of Prostate Cancer By Rathway. Onco Targets Ther. 2019;12:9143–52.
- Jiao L, Wang S, Zheng Y, Wang N, Yang B, Wang D, Yang D, Mei W, Zhao Z, Wang Z. Betulinic acid suppresses breast cancer aerobic glycolysis via caveolin-1/NF- B/c-Myc pathway. Biochem Pharmacol. 2019;161:149–62.
- Sobayashi N, Kawaguchi K, Ando A, Fujii H. Fatty acid-binding protein 5 (FABP5) promotes lipolysis of lipid droplets, de novo fatty acid (FA) synthesis and activation of nuclear factor-kappa B (NF- B) signaling in cancer cells. Biochim Biophys Acta Mol Cell Biol Lipids. 2018;1863:1057–67.
- Jiang Y, Wan Y, GW. RNA demethylase ALKBH5 promotes ovarian carcinogenesis in a simulated tumour microenvironment through stimulating NF- B pathway. J Cell Mol Med. 2020;24:6137–48.
- Siu MKY, Jiang YX, Wang JJTHY, Han CY, Tsang BK, Cheung ANY. Hexokinase 2 Res Ovarian Cancer Cell Migration, Ivasion and Stemness via FAK/ERK1/2/MMP9/NANOG/SOX9 Signaling Cascades. Cancers (Basel). 2019;11:813.
- Cheng M, Sheng L, Gao Q, Xiong Q, Zhang H, Wu M, Liang Y, Zhu F, Zhang Y, Zhang X, et al. The m(6)A methyltransfTTL3 promotes bladder cancer progression via AFF4/NF- B/MYC signaling network. Oncogene. 2019;38:3667–80.
- Li Q, Wang C, Dong W, Su Y. WTAP facilitates progression of endometrial cancer via CAV-1/NF- B axis. Cell Biol 16):1269–77.
- 59. Yin H, Zhang X, Yang P, Zhang X, Peng Y, Li D, Yu Y, Wu Y, Wang Y, Zhang J al. RNA m6A methylation orchestrates cancer gro-
- sis via macrophage reprogramming. Nat Commun. 2021;12:1394.
  Sypoxia-inducible factors in physiology and medicine. Cell. 2012;148:399–408.
- Zhou L, Wang Y, Zhou M, Zhang Y, Wang P, Li X, Yang J, Wang H. HOXA9 inhibits HIF-1 -mediated glycolysis through interacting with CRIP2 to repress cutaneous squamous cell carcinoma development. Nat Commun. 2018;9:1480.
- Feng J, Dai W, Mao Y, Wu L, Li J, Chen K, Yu Q, Kong R, Li S, Zhang J al. Simvastatin re-sensitizes hepatocellular carcinoma cells to sorafy inhibiting HIF-1 /PPAR- /PKM2-mediated glycolysis. J Exp Clin Cancer R. 2020;39:24.
- Zhang C, Samanta D, Lu H, Bullen JW, Zhang H, Chen I, He X, S GL. Hypoxia induces the breast cancer stem cell phenotype by HIFdependent and ALKBH5-mediated m6A-demethylation of NANOG mRNA. Proc Natl Acad S
- 64. Li Q, NY, Zhang L, Jiang R, Xu JYang H, Hu Y, Qiu Ju L, Tang JWang X. HIF-1 -induced expression of m6A reader YTHDF1 drives hypoxiainduced autophagy and malignancy of hepatocellular carcinoma by promoting ATG2A and ATG14 translation. Signal Transduct Target T. 2021;6:76.
- Yang N, Wang T, Li Q, Han F, Wang Z, Zhu R, Zhou Jives metabolic reprogramming in hepatocellular carcinoma cells via METTL3mediated m6A modification of HIF-1. J Cell Physiol. 2021;236:3863–80.
- Chen G, Liu B. Hypoxia induces an endometrial cancer stem-like cell phenotype via HIF-dependent demethylation of SOX2 mRNA. Oncogenesis. 2020;9:81.
- Wang Y, Yun Y, Wu B, Wen L, Wen M, Yang H, Zhao L, Liu W, Huang S, Wen N, Li Y. FOXM1 promotes reprogramming of glucose metabolism in epithelial ovarian cancer cells via activation of GLUT1 and HK2 transcription. Oncotarget. 2016;7:47985–97.
- Choi HJ, Jhe YL, K, Lim JY, L, Cheong JH. FoxM1dependent and fatty acid oxidation-mediated ROS modulation is a cellintrinsic drug resistance mechanism in cancer stem-like cells. Rx Biol. 2020;36:101589.
- Zhang S, Zhao BS, Zhou A, Lin K, Zheng S, LY, Sulman EP, Xie K, Bögler O, et al. m(6)A Demethylase ALKBH5 Maintains Tumorigenicity of Glioblastoma Stem-like Cells by Sustaining FOXM1 Expression and Cell Proliferation Program. Cancer Cell. 2017;31:591-606.e596.

- Shriwas O, Priyadarshini M, Samal SK, Rath R, Panda S, Das Majumdar SK, Muduly DK, Botlagunta M, Dash R. DDX3 modulates cisplatin resistance in OSCC through ALKBH5-mediated m(6)A-demethylation of FOXM1 and NANOG. Apoptosis. 2020;25:233–46.
- Liang Y, Hou L, Li L, Li L, Zhu L, Wang Y, Huang X, Hou Y, Zhu D, Zou H, et al. Dichloroacetate restores colorectal cancer chemosensitivity through the p53/miR-149-3p/PDK2-mediated glucose metabolic pathway. Oncogene. 2020;39:469–85.

72.

N6-Methyladenosine-Dependent Primary miR-375 Processing. Mol Ther. 2020;28:599–612.

- 111. Cheng Z, Luo C, Guo Z. LncRNA-XIST/microRNA-126 sponge mediates cell proliferation and glucose metabolism through the IRS1/PI3K/Akt pathway in glioma. J Cell Biochem. 2020;121:2170–83.
- 112. Ma JZ, Yang F, Zhou CC, Liu F, Yuan JH, Wang F, Wang TT, Xu QG, Zhou WP, Sun SH. METTL14 suppresses the metastatic potential of hepatocellular carcinoma by modulating N(6) -methyladenosine-dependent primary MicroRNA processing. Hepatology. 2017;65:529–43.
- Malakar P, Stein I, Saragovi A, Winkler R, Stern-Ginossar N, Berger M, Pikarsky E, Karni R. Long Noncoding RNA MALAT1 Regulates Cancer Glucose Metabolism by Enhancing mTOR-Mediated Translation of TCF7L2. Cancer Res. 2019;79:2480–93.
- Ye M, Dong S, Hou H, Zhang T, Shen M. Oncogenic Role of Long Noncoding RNAMALAT1 in Thyroid Cancer Progression through Regulation of the miR-204/IGF2BP2/m6A-MYC Signaling. Mol Ther Nucleic Acids. 2021;23:1–12.
- Patil DP, Chen CK, Pickering BF, Chow A, Jackson C, Guttman M, Ja rey SR. m(6)A RNA methylation promotes XIST-mediated transcriptional repression. Nature. 2016;537:369–73.
- 116. Yang X, Zhang S, He C, Xue P, Zhang L, He Z, Zang L, Feng B, Sun J, Zheng M. METTL14 suppresses proliferation and metastasis of colorectal cancer by down-regulating oncogenic long non-coding RNA XIST. Mol Cancer. 2020;19:46.
- 117. Ding C, Yi X, Chen X, Wu Z, You H, Chen X, Zhang G, Sun Y, Bu X, Wu X, et al. Warburg e ect-promoted exosomal circ\_0072083 releasing upregulates NANGO expression through multiple pathways and enhances temozolomide resistance in glioma. J Exp Clin Cancer Res. 2021;40:164.
- Li B, Zhu L, Lu C, Wang C. circNDUFB2 inhibits non-small cell lung cancer progression via destabilizing IGF2BPs and activating anti-tumor immunity. Nat Commun. 2021;12:295.
- Xu J, Wan Z, Tang M, Lin Z, Jiang S, Ji L, Gorshkov K, Mao Q, Xia S, Cen D, et al. N(6)-methyladenosine-modified CircRNA-SORE sustains sorafenib resistance in hepatocellular carcinoma by regulating -catenin signaling. Mol Cancer. 2020;19:163.
- Xue L, Li J, Lin Y, Liu D, Yang Q, Jian J, Peng J. m(6) A transferase METTL3induced IncRNA ABHD11-AS1 promotes the Warburg e ect of nonsmall-cell lung cancer. J Cell Physiol. 2021;236:2649–58.
- 121. Lu S, Han L, Hu X, Sun T, Xu D, Lİ Y, Chen Q, Yao W, He M, Wang Z, et al. N6-methyladenosine reader IMP2 stabilizes the ZFAS1/OLA1 axis and activates the Warburg e ect: implication in colorectal cancer. J Hematol Oncol. 2021;14:188.
- Wang Q, Guo X, Li L, Gao Z, Su X, Ji M, Liu J. N(6)-methyladenosine METTL3 promotes cervical cancer tumorigenesis and Warburg e ect through YTHDF1/HK2 modification. Cell Death Dis. 2020;11:911.
- 123. Yu H, Zhao K, Zeng H, Li Z, Chen K, Zhang Z, Li E, Wu Z. N(6)-methyladenosine (m(6)A) methyltransferase WTAP accelerates the Warburg e ect of gastric cancer through regulating HK2 stability. Biomed Pharmacother. 2021;133:111075.
- Shen C, Xuan B, Yan T, Ma Y, Xu P, Tian X, Zhang X, Cao Y, Ma D, Zhu X, et al. m(6)A-dependent glycolysis enhances colorectal cancer progression. Mol Cancer. 2020;19:72.
- 125. Liu XS, Yuan LL, Gao Y, Zhou LM, Yang JW, Pei ZJ. Overexpression of METTL3 associated with the metabolic status on (18)F-FDG PET/CT in patients with Esophageal Carcinoma. J Cancer. 2020;11:4851–60.
- 126. Yang D, Chang S, Li F, Ma M, Yang J, Lv X, Huangfu L, Jia C. m(6) A transferase KIAA1429-stabilized LINC00958 accelerates gastric cancer aerobic glycolysis through targeting GLUT1. IUBMB Life. 2021;73(11):1325–33.
- 127. Hou Y, Zhang Q, Pang W, Hou L, Liang Y, Han X, Luo X, Wang P, Zhang X, Li L, Meng X. YTHDC1-mediated augmentation of miR-30d in repressing pancreatic tumorigenesis via attenuation of RUNX1-induced transcriptional activation of Warburg e ect. Cell Death Di er. 2021;28:3095–124.
- 128. Yu H, Yang X, Tang J, Si S, Zhou Z, Lu J, Han J, Yuan B, Wu Q, Lu Q, Yang H. ALKBH5 Inhibited Cell Proliferation and Sensitized Bladder Cancer Cells to Cisplatin by m6A-CK2 -Mediated Glycolysis. Mol Ther Nucleic Acids. 2021;23:27–41.
- 129. Quinn JM, Greenwade MM, Palisoul ML, Opara G, Massad K, Guo L, Zhao P, Beck-Noia H, Hagemann IS, Hagemann AR, et al. Therapeutic Inhibition of the Receptor Tyrosine Kinase AXL Improves

Sensitivity to Platinum and Taxane in Ovarian Cancer. Mol Cancer Ther. 2019;18:389–98.

- Tateishi K, lafrate AJ, Ho Q, Curry WT, Batchelor TT, Flaherty KT, Onozato ML, Lelic N, Sundaram S, Cahill DP, et al. Myc-Driven Glycolysis Is a Therapeutic Target in Glioblastoma. Clin Cancer Res. 2016;22:4452–65.
- Wang Y, Lu JH, Wu QN, Jin Y, Wang DS, Chen YX, Liu J, Luo XJ, Meng Q, Pu HY, et al. LncRNA LINRIS stabilizes IGF2BP2 and promotes the aerobic glycolysis in colorectal cancer. Mol Cancer. 2019;18:174.
- 132. Yang X, Shao F, Guo D, Wang W, Wang J, Zhu R, Gao Y. WNT/ -cateninsuppressed FTO expression increases m(6)A of c-Myc mRNA to promote tumor cell glycolysis and tumorigenesis. Cell Death Dis. 2021;12:462.
- Li XD, Wang MJ, Zheng JL. Long noncoding RNA just proximal to X-inactive specific transcript facilitates aerobic glycolysis and temozolomide chemoresistance by promoting stability of PDK1 mRNA in an m6A-dependent manner in glioblastoma multiforme cells. Cancer Sci. 2021;112(11):4543–52.
- Li Z, Peng Y, Li J, Chen Z, Chen F, Tu J, Lin S. N(6)-methyladenosine regulates glycolysis of cancer cells through PDK4. Nat Commun. 2020;11:2578.
- Ou B, Liu Y, Yang X, Xu X, Yan Y, Zhang J. C5aR1-positive neutrophils promote breast cancer glycolysis through WTAP-dependent m6A methylation of ENO1. Cell Death Dis. 2021;12:737.
- Zhang C, Chen L, Liu Y, Huang J, Liu A, Xu Y, Shen Y, He H, Xu D. Downregulated METTL14 accumulates BPTF that reinforces super-enhancers and distal lung metastasis via glycolytic reprogramming in renal cell carcinoma. Theranostics. 2021;11:3676–93.
- Zaidi N, Lupien L, Kuemmerle NB, Kinlaw WB, Swinnen JV, Smans K. Lipogenesis and lipolysis: the pathways exploited by the cancer cells to acquire fatty acids. Prog Lipid Res. 2013;52:585–9.
- 138. Carracedo Á, Cantley LČ, Pandolfi PP. Cancer metabolism: fatty acid oxidation in the limelight. Nat Rev Cancer. 2013;13:227–32.
- 139. Pike LS, Smift AL, Croteau NJ, Ferrick DA, Wu M. Inhibition of fatty acid oxidation by etomoxir impairs NADPH production and increases reactive oxygen species resulting in ATP depletion and cell death in human glioblastoma cells. Biochim Biophys Acta. 2011;1807:726–34.
- 140. Svensson RU, Parker SJ, Eichner LJ, Kolar MJ, Wallace M, Brun SN, Lombardo PS, Van Nostrand JL, Hutchins A, Vera L, et al. Inhibition of acetyl-CoA carboxylase suppresses fatty acid synthesis and tumor growth of non-small-cell lung cancer in preclinical models. Nat Med. 2016;22:1108–19.
- 141. Sun D, Zhao T, Zhang Q, Wu M, Zhang Z. Fat mass and obesity-associated protein regulates lipogenesis via m(6) A modification in fatty acid synthase mRNA. Cell Biol Int. 2021;45(2):334–44.
- 142. Chen Z, Wu L, Zhou J, Lin X, Peng Y, Ge L, Chiang CM, Huang H, Wang H, He W. N6-methyladenosine-induced ERR triggers chemoresistance of cancer cells through upregulation of ABCB1 and metabolic reprogramming. Theranostics. 2020;10:3382–96.
- 143. Guo H, Wang B, Xu K, Nie L, Fu Y, Wang Z, Wang Q, Wang S, Zou X. m(6) A Reader HNRNPA2B1 Promotes Esophageal Cancer Progression via Up-Regulation of ACLY and ACC1. Front Oncol. 2020;10:553045.
- Cluntun AA, Lukey MJ, Cerione RA, Locasale JW. Glutamine Metabolism in Cancer: Understanding the Heterogeneity. Trends Cancer. 2017;3:169–80.
- 145. Li T, Le A. Glutamine Metabolism in Cancer. Adv Exp Med Biol. 2018;1063:13–32.
- Xiao Y, Thakkar KN. The m(6)A RNA demethylase FTO is a HIF-independent synthetic lethal partner with the VHL tumor suppressor. Proc Natl Acad Sci U S A. 2020;117:21441–9.
- 147. Zhu S, Wang JZ, Chen D, He YT, Meng N, Chen M, Lu RX, Chen XH, Zhang XL, Yan GR. An oncopeptide regulates m(6)A recognition by the m(6)A reader IGF2BP1 and tumorigenesis. Nat Commun. 2020;11:1685.
- Huang H, Weng H, Zhou K, Wu T, Zhao BS, Sun M, Chen Z, Deng X, Xiao G, Auer F, et al. Histone H3 trimethylation at lysine 36 guides m(6)A RNA modification co-transcriptionally. Nature. 2019;567:414–9.
- Li H, Zhang Y, Guo Y, Liu R, Yu Q, Gong L, Liu Z, Xie W, Wang C. ALKBH1 promotes lung cancer by regulating m6A RNA demethylation. Biochem Pharmacol. 2021;189:114284.
- 150. Courtney KD, Bezwada D, Mashimo T, Pichumani K, Vemireddy V, Funk AM, Wimberly J, McNeil SS, Kapur P, Lotan Y, et al. Isotope Tracing of

Human Clear Cell Renal Cell Carcinomas Demonstrates Suppressed Glucose Oxidation In Vivo. Cell Metab. 2018;28:793-800.e792.
151. Green NH, Galvan DL. MTHFD2 links RNA methylation to metabolic

- reprogramming in renal cell carcinoma. Oncogene. 2019;38:6211–25.
- 152. Li H, Song Y, He Z, Chen X, Wu X, Li X, Bai X, Liu W, Li B, Wang S, et al. Meclofenamic Acid Reduces Reactive Oxygen Species Accumulation and Apoptosis, Inhibits Excessive Autophagy, and Protects Hair Cell-Like HEI-OC1 Cells From Cisplatin-Induced Damage. Front Cell Neurosci. 2018;12:139.
- 153. Zhuang C, Zhuang C, Luo X, Huang X, Yao L, Li J, Li Y, Xiong T, Ye J, Zhang F, Gui Y. N6-methyladenosine demethylase FTO suppresses clear cell renal cell carcinoma through a novel FTO-PGC-1 signalling axis. J Cell Mol Med. 2019;23:2163–73.
- 154. Liu X, Gonzalez G, Dai X, Miao W, Yuan J, Huang M, Bade D, Li L, Sun Y, Wang Y. Adenylate Kinase 4 Modulates the Resistance of Breast Cancer Cells to Tamoxifen through an m(6)A-Based Epitranscriptomic Mechanism. Mol Ther. 2020;28:2593–604.
- 155. Shi Y, Fan S, Wu M, Zuo Z. YTHDF1 links hypoxia adaptation and nonsmall cell lung cancer progression. Nat Commun. 2019;10:4892.
- Chen P, Liu XQ, Lin X, Gao LY, Zhang S, Huang X. Targeting YTHDF1 e ectively re-sensitizes cisplatin-resistant colon cancer cells by modulating GLS-mediated glutamine metabolism. Mol Ther Oncolytics. 2021;20:228–39.
- Yan F, Al-Kali A, Zhang Z, Liu J, Pang J, Zhao N, He C. A dynamic N(6)methyladenosine methylome regulates intrinsic and acquired resistance to tyrosine kinase inhibitors. Cell Res. 2018;28:1062–76.
- Fukumoto T, Zhu H, Nacarelli T, Karakashev S, Fatkhutdinov N, Wu S. N(6)-Methylation of Adenosine of FZD10 mRNA Contributes to PARP Inhibitor Resistance. Cancer Res. 2019;79:2812–20.
- Wang L, Hui H, Agrawal K, Kang Y, Li N, Tang R. m(6) A RNA methyltransferases METTL3/14 regulate immune responses to anti-PD-1 therapy. EMBO J. 2020;39:e104514.
- 160. Huang Y, Su R, Sheng Y, Dong L, Dong Z, Xu H, Ni T, Zhang ZS, Zhang T, Li C, et al. Small-Molecule Targeting of Oncogenic FTO Demethylase in Acute Myeloid Leukemia. Cancer Cell. 2019;35:677-691.e610.
- 161. Su R, Dong L, Li Y, Gao M, Han L, Wunderlich M, Deng X, Li H, Huang Y, Gao L, et al. Targeting FTO Suppresses Cancer Stem Cell Maintenance and Immune Evasion. Cancer Cell. 2020;38:79-96.e11.
- Zhou S, Bai ZL, Xia D, Zhao ZJ, Zhao R, Wang YY. FTO regulates the chemo-radiotherapy resistance of cervical squamous cell carcinoma (CSCC) by targeting -catenin through mRNA demethylation. Mol Carcinog. 2018;57:590–7.
- Visvanathan A, Patil V, Arora A, Hegde AS, Arivazhagan A, Santosh V, Somasundaram K. Essential role of METTL3-mediated m(6)A modification in glioma stem-like cells maintenance and radioresistance. Oncogene. 2018;37:522–33.

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